



TEST FOR DIETHYLENE GLYCOL AND ETHYLENE GLYCOL IN LIQUID PREPARATIONS FOR ORAL USE

Draft proposal for inclusion in *The International Pharmacopoeia*

(27 July 2023)

DRAFT FOR COMMENTS

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For any technical queries, please contact **Dr Herbert Schmidt**, Technical Officer, Norms and Standards for Pharmaceuticals, Technical Standards and Specifications (schmidt@who.int), with a copy to Ms Sinéad Jones (jonessi@who.int, nsp@who.int).

Comments should be submitted through the online platform on or by **27 September 2023**. Please note that only comments received by this deadline will be considered for the preparation of this document.

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Test for diethylene glycol and ethylene glycol in liquid preparations for oral use

| Description | Date |
|---|--------------------|
| Text drafted by the Secretariat. | February 2023 |
| Discussion at the Consultation on Quality Control and Pharmacopoeial Specifications. | April 2023 |
| Draft text sent out for public consultation. | July – August 2023 |
| Submission to the Expert Committee on Specifications for Pharmaceutical Preparations. | October 2023 |
| Further follow-up action as required. | |

[Note from the Secretariat. Unacceptable amounts of ethylene glycol and diethylene glycol have, once again, recently been found in medicines and jeopardized the safety of patients (for more information see <https://www.who.int/teams/rethylene-glycolulation-prequalification/incidents-and-SF/full-list-of-who-medical-product-alerts> and <https://www.who.int/news/item/23-01-2023-who-urges-action-to-protect-children-from-contaminated-medicines>).

The International Pharmacopoeia (*Ph.Int.*) already prescribes a gas-chromatographic method (GC) for the test for diethylene glycol and ethylene glycol in paracetamol oral solutions. Following up on requests from WHO Member States to provide alternative procedures for National Quality Control Laboratories without access to gas chromatographs, the Secretariat proposes a tiered approach where samples are first screened for non-compliance using a semi-quantitative TLC method and suspected contaminations are subsequently confirmed by GC at regional centres.

52 *The test is proposed for inclusion in the Supplementary information section of Ph.Int.*
53 *Reference to this chapter will be made in the general monograph Liquid preparations*
54 *for oral use. The mentioned reagents, test and volumetric solutions are described in*
55 *the Reagent Section of the Ph.Int.*

56 *Users of The International Pharmacopoeia are kindly requested to provide their*
57 *feedback on the proposed tiered approach.*

58 *National Quality Control Laboratories, WHO Collaborating Centres or other*
59 *laboratories wishing to assess, demonstrate, or acquire the competency to perform the*
60 *described testing, are kindly requested to analyse a sample with a known ethylene*
61 *glycol or diethylene glycol concentration and to answer the questions listed under*
62 [*Feedback on the described procedures – Guidance for laboratories*](#) *in the Annex.*

63 *These responses will help the World Health Organization evaluate the suitability of*
64 *the described test procedures, and support regions in identifying training needs and*
65 *setting up networks/competency centres for the testing of samples for ethylene glycol*
66 *and diethylene glycol contamination.]*

Test for diethylene glycol and ethylene glycol in liquid preparations for oral use

Diethylene glycol and ethylene glycol are toxic substances used as industrial solvents and antifreeze agents that can be fatal even taken in small amounts, in particular for children.

A suitable and widely used analytical technique to test pharmaceutical products precisely and accurately for diethylene glycol and ethylene glycol is gas chromatography. For a National Quality Control Laboratory without access to a gas chromatograph, a tiered approach is described, where suspicious samples are first identified using a semi-quantitative thin-layer chromatography method (see [*Screening for non-compliance by thin-layer chromatography*](#)) and suspected contaminations subsequently confirmed at regional centres using gas chromatography (see [*Confirmatory testing by gas chromatography*](#)).

Using the thin-layer chromatography method, diethylene glycol or ethylene glycol concentrations down to 1% (m/m) are detectable. A discrimination between the two contaminants is not possible as both co-elute in the described system.

The sensitivity of the gas chromatographic method is increased by a factor of at least 10 compared to the thin-layer chromatographic procedure. Diethylene glycol or ethylene glycol concentrations down to 0.1% (m/m) for each contaminant can be determined.

Laboratories whose investigations have confirmed diethylene glycol or ethylene glycol contaminations in excipients or finished products shall inform the responsible regulatory authorities without delay.

92 **Screening for non-compliance by thin-layer chromatography**

93 Carry out the test as described under *1.14.1 Chromatography*, Thin-layer
94 chromatography, using a plate coated with silica gel R5 and of a suitable size (e.g. 10
95 cm x 10 cm, 10 cm x 20 cm or 20 cm x 20 cm).

96 Saturate a chromatographic tank of a suitable size by lining its walls with filter paper.
97 Pour a sufficient quantity of a freshly prepared mixture of toluene R, acetone R and
98 ammonia (~100 g/L) TS (5:85:10 V/V/V) into the chromatographic tank. After
99 impregnation of the filter paper, the mobile phase shall form a layer of appropriate
100 depth, usually 1 cm. Replace the lid and allow the tank to stand for 1 hour at 20-25 °C
101 to complete the saturation of the tank.

102 Prepare the following solutions, using methanol R as a solvent.

103 For solution (A) (*sample solution*), transfer 1.000 g of the oral solution under
104 investigation to a 5.0 mL volumetric flask and fill up to volume.

105 For solution (B) (*diethylene glycol reference solution, 20% (m/m), 40 mg diethylene*
106 *glycol/mL*), dissolve 1.000 g of diethylene glycol R in 25.0 mL.

107 For solution (C) (*diethylene glycol reference solution, 10% (m/m), 20 mg diethylene*
108 *glycol /mL*), dilute 10.0 mL of solution (B) to 20.0 mL.

109 For solution (D) (*diethylene glycol reference solution, 1% (m/m), 2 mg diethylene*
110 *glycol /mL*), dilute 5.0 mL of solution (B) to 100.0 mL.

111 For solution (E) (*ethylene glycol reference solution, 20% (m/m), 40 mg ethylene*
112 *glycol/mL*), dissolve 1.000 g of ethylene glycol R in 25.0 mL.

113 For solution (F) (*ethylene glycol reference solution, 10% (m/m), 20 mg ethylene*
114 *glycol /mL*), dilute 10.0 mL of solution (E) to 20.0 mL.

115 For solution (G) (*ethylene glycol reference solution, 1% (m/m), 2 mg ethylene glycol*
116 */mL*), dilute 5.0 mL of solution (E) to 100.0 mL.

117 Apply separately to the plate, 2 μ L of each of solutions (A), (B), (C), (D), (E), (F) and
118 (G) at a distance of about 2 cm from the lower edge and from the sides of the plate
119 and on a line parallel to the lower edge. Allowing the standard solutions to bracket the
120 sample solution. Use a device that allows the reproducible dispensing of the
121 prescribed volume onto the plate (e.g. micro capillary pipettes).

122 Apply the solutions in sufficiently small portions to obtain circular spots of 2-5 mm in
123 diameter. Allow an interval of at least 10 mm between the centres of circular spots.

124 When the solvent of the applied solutions has evaporated, place the plate in the
125 chromatographic tank, ensuring that the plate is as vertical as possible and that the
126 spots are above the surface of the mobile phase. Close the chromatographic tank and
127 maintain it at 20-25 °C, protected from sunlight and placed in a draught-free area.

128 Remove the plate when the mobile phase has moved over a distance of about 7 cm
129 (for 10 cm x 10 cm or 10 cm x 20 cm plates) or 15 cm (for 20 cm x 20 cm plates)
130 measured between the points of application and the solvent front.

131 Dry the plate and visualise the spots by spraying the plate with a solution containing
132 0.632 g of potassium permanganate R in 100 mL of acetone R. Heat the plate at
133 105 °C for 15 minutes. Ethylene glycol and diethylene glycol appear as yellow spots
134 on a reddish background.

135 In the chromatograms obtained, the following substances are eluted with the following
136 R_f values: sorbitol about 0.1; glycerol about 0.45; diethylene glycol and ethylene
137 glycol about 0.55; propylene glycol about 0.65.

138 The test is not valid unless the chromatograms, obtained with solutions (C) and (F),
139 show spots due to diethylene glycol or ethylene glycol with a similar position and
140 appearance (diethylene glycol and ethylene glycol co-elute).

141 Identify the spots due to diethylene glycol or ethylene glycol in the chromatograms
142 obtained and estimate their percentage content (m/m) in the liquid preparation of oral
143 use under investigation by comparing the intensity of the spot due to diethylene glycol

or ethylene glycol in the chromatogram obtained with solution (A), if present, with the intensity of the spots due to diethylene glycol or ethylene glycol in the chromatograms obtained with solutions (B) to (G).

If a contamination of the oral solution under investigation is determined, confirm the estimated diethylene glycol or ethylene glycol concentration by gas chromatography.

Confirmatory testing by gas chromatography

Carry out the test as described under [1.14.1 Chromatography, Gas chromatography](#) using the internal standard method.

For the procedure, use a capillary glass or quartz column (30 m × 0.53 mm), the inner surface of which is coated with a thick layer of macrogol 20M R (1.0 µm). Maintain the temperature of the column at 100 °C for 5 minutes. Increase the temperature at a rate of 10 °C per minute to 230 °C and maintain it at this point for 4 minutes. Maintain the temperature of the injection port and the detector at 230 °C. Use helium R as the carrier gas with a linear velocity of about 38 cm per second. For application of the solutions, use splitless injection followed by a split ratio of 1:20 after 30 seconds. Use a flame-ionization detector for detection.

Prepare the following solutions in a 1:1 mixture of acetonitrile R and water R.

For solution (IS), weigh 0.175 g of the internal standard 1,3-butanediol R and dilute to 100.0 mL.

For solution (A), weigh 10.000 g of the liquid preparation for oral solution under investigation, add 4.0 mL of solution (IS) and dilute to 100.0 mL.

For solution (B), weigh 10.000 g of oral solution under investigation and dilute to 100.0 mL.

For solution (C), weigh 0.500 g of each ethylene glycol RS, propylene glycol R and diethylene glycol RS and dilute to 50.0 mL.

169 For solution (D) mix 25.0 mL of solution (C) with 2.0 mL of solution (IS) and dilute
170 to 50.0 mL.

171 For solution (E) mix 10.0 mL of solution (C) with 4.0 mL of solution (IS) and dilute
172 to 100.0 mL.

173 For solution (F) mix 1.0 mL of solution (C) with 4.0 mL of solution (IS) and dilute to
174 100.0 mL.

175 For solution (G) dilute 2.0 mL of solution (C) to 20.0 mL. Mix 2.0 mL of this solution
176 with 4.0 mL solution (IS) and dilute to 100 mL.

177 Inject separately 1.0 μ l each of solutions (A), (B), (C), (F) and (G) and record the
178 chromatograms.

179 Measure the response of the peaks corresponding to diethylene glycol, ethylene
180 glycol, propylene glycol and 1,3-butanediol in the chromatograms obtained.

181 In the chromatogram obtained with solution (E), the analyte peaks are eluted at the
182 following relative retention with reference to 1,3-butanediol (retention time about 14
183 minutes): propylene glycol about 0.73; ethylene glycol about 0.76; diethylene glycol
184 about 1.04; and glycerol about 1.27.

185 The test is not valid unless the resolution, between the peaks corresponding to
186 ethylene glycol and propylene glycol in the chromatogram obtained with solution (E),
187 is at least 4.0 and no peak having the same retention time as 1,3-butanediol can be
188 detected in the chromatogram obtained with solution (B). Also, the test is not valid
189 unless, in the chromatogram obtained with solution (G), the signal-to-noise ratios of
190 the peaks due to diethylene glycol and ethylene glycol are at least 10.

191 Calculate the ratios between the responses of the peaks due to diethylene glycol/
192 ethylene glycol and the responses of the peak due to 1,3 butanediol in the
193 chromatograms obtained with the solutions (A), (D), (E) and (F).

194 Prepare a calibration function each for ethylene glycol and diethylene glycol by
195 plotting the ratios obtained for solutions (D), (E) and (F) on the ordinate against the
196 amount of the reference standard on the abscissa. Consider the declared content of
197 $C_2H_6O_2$ or $C_4H_{10}O_3$ in ethylene glycol RS and diethylene glycol RS

198 Use the ratio obtained for solution (A) and the calibration functions to calculate the
199 percentage content (m/m) of diethylene glycol and ethylene glycol in the liquid
200 preparation for oral use. The percentage concentration (m/m) of diethylene glycol or
201 ethylene glycol in the liquid preparation for oral use should be below 0.1%.

202 If the result obtained is above 5.0%, prepare solution (1) using a lower weight of
203 liquid preparation for oral use under investigation and repeat the analysis.

204 **International Chemical Reference Substances to be established**

205 Ethylene glycol ICRS

206 Diethylene glycol ICRS

207 **Reagents to be added to The International Pharmacopoeia:**

208 **1,3-Butanediol R**

209 $C_4H_{10}O_2$; CAS Reg. No. 107-88-0.

210 Contains not less than 99.0% of $C_4H_{10}O_2$.

211 **Ethylene glycol R**

212 Ethane-1,2-diol; $C_2H_6O_2$; CAS Reg. No. 107-21-1.

213 Contains not less than 99.0% of $C_2H_6O_2$.

214 *Description.* Colourless, slightly viscous liquid, hygroscopic, miscible with water R
215 and dehydrated ethanol R.

216 *Relative density.* $d_{20}^{20} = 1.113$ to 1.115.

217 *Refraction index.* n_D^{20} about 1.432.

218 *Boiling point.* About 198 °C.

219 *Melting point.* About -12 °C.

220 *Acidity.* To 10 mL, add 20 mL of water R and 1 mL of phenolphthalein/ethanol TS.
221 Not more than 0.15 mL of sodium hydroxide (0.02 mol/L) VS is required to change
222 the colour of the indicator to pink.
223 Water (2.8): Not more than 2 mg/g.

224 **Diethylene glycol R**

225 2,2'-Oxydiethanol; $C_4H_{10}O_3$; CAS Reg. No. 111-46-6.
226 Contains not less than 99.5% of $C_4H_{10}O_3$.
227 *Description.* Clear, colourless liquid, hygroscopic, miscible with water R, acetone R
228 and dehydrated ethanol R.
229 *Relative density.* d_{20}^{20} about 1.118.
230 *Refraction index.* n_D^{20} about 1.447.
231 *Boiling point.* 244 °C to 246 °C.
232 *Storage.* In an airtight container.

233

234

235 Annex:

236 **Feedback on described analytical procedures –**
237 **Guidance for laboratories**

238

239 National Quality Control Laboratories, WHO Collaborating Centres or other
240 laboratories wishing to assess, demonstrate or acquire the competency to perform the
241 described testing are kindly requested to:

- 242 I. analyse a sample with a **known** ethylene glycol or diethylene glycol
243 concentration according to the provision *Test for diethylene glycol and ethylene*
244 *glycol in liquid preparations for oral use*; and
245 II. answer the questions below and send the responses, together with the requested
246 documents, to schmidth@who.int.

247 These responses will help the World Health Organization evaluate the suitability of
248 the described test procedure and support regions in identifying training needs and
249 setting up networks/competency centres for the testing of samples for ethylene glycol
250 and diethylene glycol contamination.

251 **To I.**

252 To be able to evaluate the performance of the test procedure, the laboratory is
253 requested to test a sample with a **known** ethylene glycol or diethylene glycol
254 concentration. Such a sample can be prepared by adding a known ethylene glycol
255 or diethylene glycol concentration to a sample free of these substances from the
256 market.

257 Please perform the following steps:

258 Take a liquid preparation for oral use from your market and ensure that it is not
259 contaminated with diethylene glycol or ethylene glycol by analysing it according to

the provision: [*Test for diethylene glycol and ethylene glycol in liquid preparations for oral use, Confirmation by gas chromatography.*](#)

Spike the preparation with either ethylene glycol or diethylene glycol, accurately weighed, so that the sample contains between 1.0% to 15.0% (m/m) of one of these contaminants.

Test the spiked sample firstly according to the analytical procedure [*Screening for non-compliance by thin-layer chromatography*](#) and secondly, if a gas chromatograph is available, according to [*Confirmatory testing by gas chromatography*](#). As the International Chemical Reference Substances on diethylene glycol and ethylene glycol are currently under establishment, use other suitable reference substances for the latter procedure.

To II.

Please summarize the results of your investigations in a comprehensive laboratory report. Respond, in particular, to the following questions/points:

1. Name and address of your laboratory.
2. Name, title and email address of a contact person.
3. Which sample (name, manufacturer, lot number) has been spiked with diethylene glycol or ethylene glycol? Please provide details of the composition of the sample (active ingredient(s), strength(s) and excipients).
4. Which of the tests were performed: *Screening for non-compliance by thin-layer chromatography*, *Confirmatory testing by gas chromatography*, or both? If Confirmatory testing by gas chromatography was not performed, please provide an explanation as to why.
5. While performing the *Screening for non-compliance by TLC*, did the analyst deviate from the prescribed procedure? If yes, please describe the deviations in detail and provide a rational as to why changes were made.

- 286 6. While performing the *Confirmatory testing by gas chromatography*, did the
287 analyst deviate from the prescribed procedure? If yes, please describe the
288 deviations in detail and provide a rational as to why changes were made.
- 289 7. What was the diethylene glycol or ethylene glycol target concentration in the
290 spiked sample (calculated based on the amount with which the sample was
291 spiked)?
- 292 8. Which diethylene glycol or ethylene glycol concentration/percentage content
293 was estimated based on the performance of the *Screening of non-compliance*
294 *by TLC* on the spiked sample?
- 295 9. Which diethylene glycol or ethylene glycol concentration/percentage content
296 was determined based on the performance of the *Confirmatory testing by gas*
297 *chromatography* on the spiked sample?
- 298 10. Please include (photocopies of) the chromatogram(s) obtained when
299 performing the TLC and GC methods. Indicate on the print-outs which
300 solutions were analysed.
- 301 11. Did the analyst encounter problems/challenges when performing the analysis?
302 If so, please describe them in detail.
- 303 12. Please evaluate your laboratory's competency to perform the *Screening of*
304 *non-compliance by TLC* on liquid preparations for oral use. If your laboratory
305 does not yet have this competency, please identify/describe areas/topics for
306 which your laboratory would need further training/guidance in order to obtain
307 this competency.
- 308 13. Please evaluate your laboratory's capacity to perform the *Confirmatory*
309 *testing by GC*. If your laboratory does not yet have this competency, please
310 identify/describe areas/topics for which your laboratory would need further
311 training/guidance in order to obtain this competency.

312 ***